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# Beta 3-adrenoceptor stimulation induces vasorelaxation mediated essentially by endothelium-derived nitric oxide in rat thoracic aorta

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- 1 The relaxant effects of isoprenaline may result from activation of another  $\beta$ -adrenoceptor subtype in addition to  $\beta_1$  and  $\beta_2$ . This study evaluated the role of a third  $\beta$ -adrenoceptor subtype,  $\beta_3$ , in  $\beta$ -adrenoceptor-induced relaxation of rat thoracic aorta by isoprenaline.
- 2 Isoprenaline produced a concentration-dependent relaxation of phenylephrine pre-contracted rings of the thoracic aorta (pD $_2$ =7.46 $\pm$ 0.15;  $E_{max}$ =85.9 $\pm$ 3.4%), which was partially attenuated by endothelium removal ( $E_{max}$ =66.5 $\pm$ 6.3%) and administration of the nitric oxide (NO) synthase inhibitor, L-NG-monomethyl arginine (L-NMMA) ( $E_{max}$ =61.3 $\pm$ 7.9%).
- 3 In the presence of nadolol, a  $\beta_1$  and  $\beta_2$ -adrenoceptor antagonist, isoprenaline-induced relaxation persisted (E<sub>max</sub> = 55.6 ± 5.3%), but occurred at higher concentrations (pD<sub>2</sub> = 6.71 ± 0.10) than in the absence of nadolol and lasted longer.
- 4 Similar relaxant effects were obtained with two  $\beta_3$ -adrenoceptor agonists: SR 58611 (a preferential  $\beta_3$ -adrenoceptor agonist), and CGP 12177 (a partial  $\beta_3$ -adrenoceptor with  $\beta_1$  and  $\beta_2$ -adrenoceptor antagonistic properties). SR 58611 caused concentration-dependent relaxation (pD<sub>2</sub>=5.24±0.07; E<sub>max</sub>=59.5±3.7%), which was not modified by pre-treatment with nadolol but antagonized by SR 59230A, a  $\beta_3$ -adrenoceptor antagonist. The relaxation induced by SR 58611 was associated with a 1.7 fold increase in tissue cyclic GMP content.
- 5 Both relaxation and the cyclic GMP increase induced by SR 58611 were greatly reduced by endothelium removal and in the presence of L-NMMA.
- 6 We conclude that in the rat thoracic aorta,  $\beta_3$ -adrenoceptors are mainly located on endothelial cells, and act in conjuction with  $\beta_1$  and  $\beta_2$ -adrenoceptors to mediate relaxation through activation of an NO synthase pathway and subsequent increase in cyclic GMP levels.

**Keywords:** Rat thoracic aorta; relaxation;  $\beta_3$ -adrenoceptor; endothelium; nitric oxide; cyclic GMP

Abbreviations

E<sub>max</sub>, maximal relaxant response; CGP 12177, 4-[3-*t*-butylamino-2-hydroxypropoxy]benzimadazol-2-one; IBMX, 3-isobutyl-1-methylxanthine; L-NMMA, L-N<sup>G</sup>-monomethyl-arginine; SR 58611, (RS)-*N*-[(25)-7-ethoxycarbonyl-methoxy-1,2,3,4-tetrahydronaphth-2-yl]-(2R)-2-(3-chlorophenyl)-2 hydroethanamine hydrochloride; SR 59230A, 3-(2-ethylphenoxy)-1-[(1S)1,2,3,4-tetrahydronaphth-1-ylaminol]-(2S)-2-propanol oxalate

## Introduction

The cloning of a third  $\beta$ -adrenoceptor subtype ( $\beta_3$ ) in 1989 (Emorine *et al.*, 1989) has allowed an explanation of some of the effects of catecholamines which are not related to simultaneous or concomitant activation of  $\beta_1$ - and  $\beta_2$ -adrenoceptors.  $\beta_3$ -adrenoceptors were subsequently found to mediate lipolysis in adipose tissues (for review, see Lafontan, 1994) and the relaxation of gastrointestinal (for review, see Manara *et al.*, 1995b) and airway (for review, see Martin & Advenier, 1995) smooth muscle. More recently,  $\beta_3$ -adrenoceptors have been characterized in human heart, activation of which induces a negative inotropic effect (Gauthier *et al.*, 1996)

In vascular smooth muscle,  $\beta$ -adrenoceptors were initially classified as  $\beta_2$ -adrenoceptors (Lands *et al.*, 1967). Later studies using more selective agonists and antagonists showed that vascular relaxation could result from activation of either  $\beta_1$ - or  $\beta_2$ -adrenoceptor subtypes and that the involvement of

each subtype depended on the vascular bed and the species studied. In most vessels, the relaxation induced by isoprenaline is mediated essentially through the activation of  $\beta_2$ -adrenoceptors, with little contribution from  $\beta_1$ -adrenoceptors. The involvement of a third  $\beta$ -adrenoceptor subtype in  $\beta$ adrenoceptor agonist-induced vasorelaxation has been suggested in several studies. Pindolol, a non-specific  $\beta$ -adrenoceptor antagonist with significant intrinsic sympathomimetic activity, induced the relaxation of canine isolated perfused mesenteric vessels (Clark & Bertholet, 1983) and rat aorta (Doggrell, 1990). In rat aorta, propranolol, a non-selective  $\beta$ adrenoceptor antagonist, inhibited the vasorelaxant effect of isoprenaline, a non-selective  $\beta$ -adrenoceptor antagonist, inhibited the vasorelaxant effect of isoprenaline at higher concentrations than those expected solely from activation of  $\beta_1$ - and  $\beta_2$ -adrenoceptors. This effect was attributed to catecholamine-induced stimulation of atypical  $\beta$ -adrenoceptors (Oriowo, 1995) as well as  $\beta_2$ -adrenoceptors and a small population of  $\beta_1$ -adrenoceptors (O'Donnell & Wanstall, 1985). The possibility of an atypical  $\beta$ -adrenoceptor in vessels was strengthened by the use of preferential  $\beta_3$ -adrenoceptor agonists (BRL 37344, CL 316243). In in vivo studies, these agents produced vasodilatation in dogs (Tavernier et al., 1992;

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Shen et al., 1994; 1996) and to a lesser extent in rats (Shen et al., 1996).

The main purpose of the present study was to determine the role of  $\beta_3$ -adrenoceptors in relaxation of the rat thoracic aorta induced by  $\beta$ -adrenoceptor agonists. The effects of two  $\beta_3$ adrenoceptor agonists, SR 58611 (a preferential agonist) and CGP 12177 (a partial agonist which also possesses  $\beta_1$ - and  $\beta_2$ adrenoceptor antagonistic properties (Blin et al., 1993)), were compared with those of isoprenaline in the absence and presence of a  $\beta_1$ - and  $\beta_2$ -adrenoceptor-blocking drug, nadolol (Lee et al., 1975). A second purpose of this study was to characterize the cellular coupling pathway involved after stimulation of vascular  $\beta_3$ -adrenoceptors, especially the role of the endothelium and cyclic GMP-dependent and/or -independent NO pathways.

#### Methods

Tissue preparation and tension studies in rat aortic rings

Adult male Wistar rats (250-300 g) were anaesthetized with pentobarbital (30 mg kg<sup>-1</sup> i.p.). Descending thoracic aortae were isolated, cleared of fat and connective tissue and cut into 3 mm rings. In some rings, the endothelium was removed by gentle rubbing of the intimal surface with a fine pair of small forceps. Rings were suspended on stainless-steel wires in a 20 ml organ bath containing Krebs solution composed as follows (mm): NaCl, 118.3; KCl, 4.7; MgSO<sub>4</sub>, 1.2; KH<sub>2</sub>PO<sub>4</sub>, 1.2; NaHCO<sub>3</sub>, 25; EDTA (ethylenediaminetetraacetic acid), 0.016; glucose, 11.1; and CaCl<sub>2</sub>, 2.5 (pH 7.4). Bath temperature was maintained at 37°C, and the Krebs solution was continuously oxygenated with a 95% O<sub>2</sub>, 5% CO<sub>2</sub> gas mixture. Rings were progressively stretched to a resting tension of 2 g. Isometric tension was recorded by a force displacement transducer (IT2, EMKA Technologies, Paris, France) and displayed on a computer (IOX1.5.7 software, EMKA Technologies). Data were analysed using Datanalyst software (EMKA Technologies).

Functional endothelium was checked by the presence of at least 70% relaxation in response to acetylcholine (1  $\mu$ M) in rings pre-contracted with phenylephrine (0.3  $\mu$ M). In denuded vascular rings, endothelium removal was confirmed by the absence of acetylcholine-induced relaxation. Some rings were equilibrated in Krebs containing nadolol (a  $\beta_1$ and  $\beta_2$ -adrenoceptor antagonist), SR 59230A (a  $\beta_3$ -adrenoceptor antagonist), or the NO synthase inhibitor, NGmonomethyl-L-arginine monoacetate (L-NMMA), 30 min. Control rings were not treated during this period. Aortic rings were contracted again with phenylephrine to obtain a similar magnitude of sustained tension as in control rings. A cumulative concentration-response curve to either isoprenaline or a  $\beta_3$ -adrenoceptor agonist was then constructed. The concentration of phenylephrine  $(0.1-1 \mu M)$ was adjusted to produce a similar level of tone for each experimental condition. Relaxation produced by each concentration of  $\beta$ -adrenoceptor agonist was measured after steady-state was reached. Values are expressed as the percentage change in the maximal tension of vessel rings after addition of phenylephrine.

As SR 58611 and CGP 12177 induced long-lasting relaxation, spontaneous time-dependent relaxation was evaluated in control vessels. This phenomenon was taken into account by systematic subtraction of the corresponding spontaneous relaxation obtained in control vessels from the relaxation produced by the  $\beta_3$ -adrenoceptor agonist.

Cyclic GMP assay

Frozen aortic rings were individually ground in a pestle and mortar with ice-cold 6% v  $v^{-1}$  trichloroacetic acid plus 100  $\mu$ M 3-isobutyl-1-methylxanthine (IBMX). After centrifugation at  $1500 \times g$  for 10 min at 4°C, trichloroacetic acid was extracted by washing supernatants three times with water-saturated ether (five volumes of ether to one volume of supernatant). The remaining ether was evaporated by heating the samples to 70°C for 5 min. Cyclic GMP contents were measured using an enzyme immunoassay kit (Cayman Chemical Company, Ann Arbor, MI, U.S.A.). Absorbance was read on a spectrophotometer at 405 nm. A standard curve was drawn for each assay. The mean value was calculated from duplicate measurements of each sample and normalized to total cell protein content as determined by the method of Lowry et al. (1951).

#### Drugs

L-phenylephrine hydrochloride, acetylcholine chloride, (-)isoprenaline, nadolol, IBMX and prostaglandin  $F_{2\alpha}$  Tris salt  $(PGF_{2\alpha})$  were obtained from the Sigma Chemical Co. (St. Louis, MO, U.S.A.). L-NMMA was purchased from Calbiochem (La Jolla, CA, U.S.A.) and CGP 12177 (4-[3-tbutylamino-2-hydroxypropoxy]benzimidazol-2-one) from RBI (Natick, MA, U.S.A.). SR 58611 [(RS)-N-[(25)-7-ethoxycarbonylmethoxy-1,2,3,4-tetrahydronaphth-2-yl]-(2R)-2-(3-chlorophenyl)-2 hydroethanamine hydrochloridel and SR 59230A (3-(2-ethylphenoxy)-1-[(1S)1,2,3,4-tetrahydronaphth-1-ylaminol]-(2S)-2-propanol oxalate) were a generous gift from Sanofi Recherche (Montpellier, France). All drugs were prepared as stock solutions in distilled water, with the exception of nadolol which was dissolved in hydrochloric acid before being neutralized to pH 7.4, and SR 59230 which was dissolved in dimethylsulphoxide (DMSO; Sigma Chemical) such that the final concentration of the solvent in the organ bath was less than 0.1% v v<sup>-1</sup>. At this concentration, the solvent alone had no effect on the tissue.

## Data and statistical analysis

Results are expressed as the mean  $\pm$  s.e.mean of *n* experiments. The statistical significance of a drug effect was assessed using one-way analysis of variance (ANOVA) followed by a Dunnett's test. Comparison of the different concentration response curves was performed by two-way ANOVA. To determine agonist potencies from concentration response curves, concentrations producing 50% of maximum effect (EC<sub>50</sub>) were calculated by fitting curves with the Boltzmann equation. pD2 values were then determined according to the equation  $pD_2 = -\log(\text{molar } EC_{50})$  and compared using Student's *t*-test (P < 0.05 being considered as significant).

#### Results

Phenylephrine  $(0.3 \mu M)$  increased the resting tone of rat aortic rings when endothelium was intact. As the contractile response to 0.3 µM phenylephrine was more marked when endothelium was removed or L-NMMA was present, the concentration of phenylephrine was decreased to  $0.1 \, \mu M$  to obtain a tone level equivalent to that in control rings. In this way, the contractile tone caused by phenylephrine was similar in the different experimental conditions shown in Table 1. When the endothelium was intact, acetylcholine  $(1 \mu M)$  induced a significant relaxation (up to 70%), whereas no relaxant effect was observed in endothelium-denuded rings (data not shown).

#### Isoprenaline-induced relaxation

As shown in Figures 1A and 2, isoprenaline  $(0.001-3~\mu\text{M})$  induced concentration—dependent relaxation in control aortic rings (with an intact endothelium) pre-contracted with phenylephrine (pD<sub>2</sub> value of  $7.46\pm0.15~(n=10)$  and an E<sub>max</sub> of  $85.9\pm3.4\%~(n=10)$ ). This effect was produced rapidly, reaching its maximum level within about 4 min after isoprenaline administration. In rings without endothelium, the concentration-relaxation curve for isoprenaline was shifted to the right (pD<sub>2</sub>= $6.50\pm0.11$ ), and the maximal relaxant effect was reduced (E<sub>max</sub>= $66.5\pm6.3\%$ ; n=7; P<0.05~versus isoprenaline in intact rings; Figure 2).

# Effect of nadolol on isoprenaline-induced relaxation

In another set of experiments, the effect of isoprenaline was tested after administration of nadolol for 30 min. Nadolol (10  $\mu$ M) alone failed to produce any intrinsic effect on

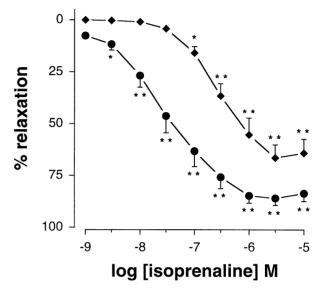
**Table 1** Effect of endothelium-removal, L-NMMA and nadolol on contractile responses induced by phenylephrine in rat thoracic aortic rings

	Tension (g)	Number of experiments
Control rings	$3.93 \pm 0.14$	19
Denuded rings	$4.29 \pm 0.11$	14
L-NMMA (100 μm)*	$4.19 \pm 0.17$	18
Nadolol (10 $\mu$ M)*	$4.03 \pm 0.17$	16

Data are expressed as changes in isometric tension (g) and shown as mean  $\pm$  s.e.mean for n experiments. \*Changes in contractile responses induced by phenylephrine was measured 30 min after administration of L-NMMA or nadolol.

phenylephrine-induced concentration of aortic rings. The relaxant effect of isoprenaline was not abolished in the presence of nadolol. However, the effect occurred at higher concentrations of isoprenaline (up to  $0.03~\mu M$ ), and relaxation was both delayed and long-lasting (Figure 1B). The maximal relaxation for a given concentration was observed within 10-

- Isoprenaline
- ♦ Isoprenaline + endothelium removal



**Figure 2** Concentration-response curves to isoprenaline in rat thoracic aortic rings with (n=10) or without endothelium (n=7). Results are expressed as the percentage of relaxation from the maximal level of contraction induced by phenylephrine. Each point is the mean of n experiments, and vertical lines show the s.e.mean. When no error bar is shown, the error is smaller than the symbol. \*P < 0.05 and \*\*P < 0.01 indicate significant differences from control.

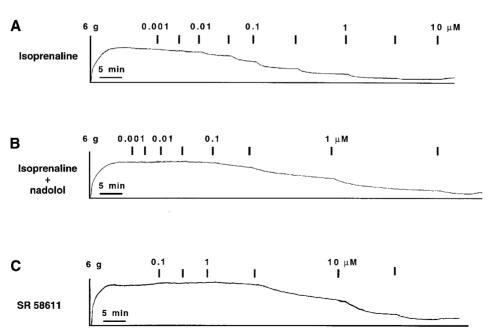
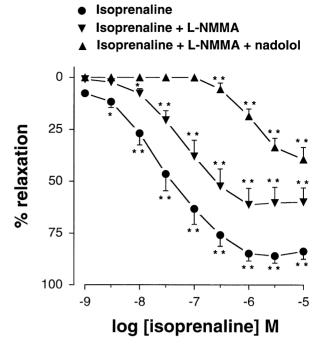


Figure 1 Typical recordings of relaxant effects of  $\beta$ -adrenoceptor agonists in rat thoracic aortic rings constricted with phenylephrine (0.3 μM). (A) isoprenaline induces a rapid and potent relaxant effect at concentrations from 0.001–1 μM. (B) In the presence of 10 μM nadolol, a  $\beta_1$ - and  $\beta_2$ -adrenoceptor antagonist, isoprenaline still induces relaxation (characterized by its long duration) at concentrations up to 0.03 μM. (C) SR 58611, a  $\beta_3$ -adrenoceptor agonist, induces a concentration-dependent relaxant effect similar to that exhibited by isoprenaline in the presence of nadolol.

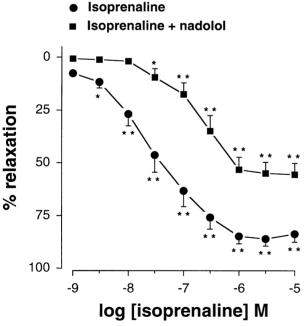
12 min after administration of isoprenaline. This long-lasting effect led us to evaluate spontaneous time-dependent relaxation of vessels in control rings pre-treated with nadolol (10  $\mu$ M), which was found to be responsible for 17.6±1.1% of relaxant effect at the end of the experiment (about 90 min after administration of phenylephrine; n=5). The corresponding spontaneous relaxation of control vessels was then subtracted from that exhibited by isoprenaline plus nadolol (10  $\mu$ M). In these conditions, the concentration response curve to isoprenaline was shifted to the right in the presence of nadolol (10  $\mu$ M) (pD<sub>2</sub>=6.71±0.10; n=9; P<0.01 versus isoprenaline alone), and the maximal relaxant effect of isoprenaline was significantly reduced ( $E_{max}=55.6\pm5.3\%$ ; n=9; P<0.01 versus isoprenaline alone; Figure 3).

# Effects of L-NMMA on isoprenaline-induced relaxation

To characterize the involvement of a NO synthase pathway in the relaxant effect of  $\beta$ -adrenoceptor stimulation, the ability of the NOS inhibitor, L-NMMA to alter the response to isoprenaline was determined. L-NMMA shifted the relaxant response to isoprenaline to the right (pD<sub>2</sub>=6.57±0.10; n=9; P<0.01 versus isoprenaline alone) and decreased the maximal level of relaxation from  $85.9\pm3.4\%$  to  $61.3\pm7.9\%$  (n=9; P<0.01 versus isoprenaline alone; Figure 4). The concentration-relaxant response curve to isoprenaline was shifted toward higher concentrations in the presence of L-NMMA (100  $\mu$ M) plus nadolol (10  $\mu$ M) (pD<sub>2</sub>=5.81±0.20; n=5; P<0.01 versus isoprenaline alone and P<0.05 versus isoprenaline plus nadolol) than in the presence of either agent alone. In these conditions, the maximal relaxant effect of isoprenaline



**Figure 4** Concentration-response curves for isoprenaline in the absence (n=10) and presence of L-NMMA  $(100 \ \mu \text{M}; n=9)$  and in the presence of nadolol  $(10 \ \mu \text{M}) + \text{L-NMMA} \ (n=5)$  in rat thoracic aortic rings pre-contracted with phenylephrine. Results are expressed as the percentage of relaxation from the maximal contraction level induced by phenylephrine. Each point is the mean of n experiments, and vertical lines show the s.e.mean. When no error bar is shown, the error is smaller than the symbol. \*P < 0.05 and \*\*P < 0.01 indicate significant differences from control.



**Figure 3** Concentration-response curves for isoprenaline in the absence (n=10) and presence of nadolol  $(10~\mu\mathrm{M}; n=9)$  in rat thoracic aortic rings pre-contracted with phenylephrine. As the relaxant effect of isoprenaline in the presence of nadolol was slow in reaching steady-state, spontaneous relaxation of control vessels was subtracted from the relaxation exhibited by isoprenaline in the presence of nadolol. Results are expressed as the percentage of relaxation from the maximal contraction level induced by phenylephrine. Each point is the mean of n experiments, and vertical lines show the s.e.mean. When no error bar is shown, the error is smaller than the symbol. \*P < 0.05 and \*\*P < 0.01 indicate significant differences from control.

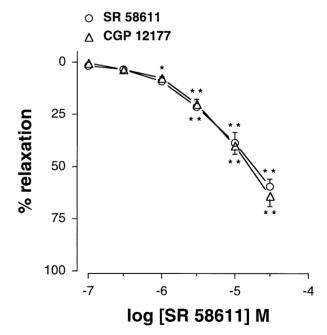
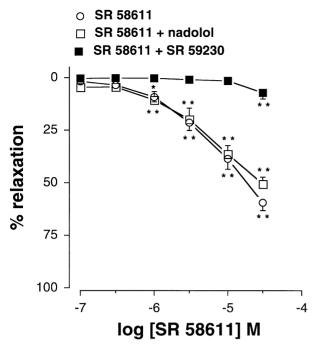
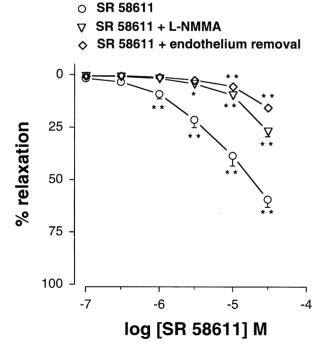


Figure 5 Concentration-response curves for SR 58611, a preferential  $\beta_3$ -adrenoceptor agonist (n=11), and CGP 12177, a partial  $\beta_3$ -adrenoceptor agonist (n=6), in rat thoracic aortic rings constricted with phenylephrine. The mean curves resulting from subtraction of the spontaneous relaxation of control vessels are shown. Results are expressed as the percentage of relaxation from the maximal contraction level induced by phenylephrine. Each point is the mean of n experiments, and vertical lines show the s.e.mean. When no error bar is shown, the error is smaller than the symbol. \*P<0.05 and \*\*P<0.01 indicate significant differences from control.



**Figure 6** Concentration-response curves for SR 58611 in the absence (n=11) and presence of nadolol  $(10 \ \mu \text{M}; n=7)$  or SR 59230A  $(10 \ \mu \text{M}; n=6)$  in rat thoracic aortic rings constricted with phenylephrine. The mean curves resulting from subtraction of the spontaneous relaxation of control vessels pre-treated or not with nadolol or SR 59230A are shown. Results are expressed as the percentage of relaxation from the maximal contraction level induced by phenylephrine. Each point is the mean of n experiments, and vertical lines show the s.e.mean. When no error bar is shown, the error is smaller than the symbol. \*P < 0.05 and \*P < 0.01 indicate significant differences from control.



**Figure 7** Concentration-response curves for SR 58611 in rat thoracic aortic rings with (n=11) or without endothelium (n=7) and after pre-treatment with L-NMMA (100  $\mu$ M; n=9). Results are expressed as the percentage of relaxation from the maximal contraction level induced by phenylephrine. Each point is the mean of n experiments, and vertical lines show the s.e.mean. When no error bar is shown, the error is smaller than the symbol. \*P<0.05 and \*\*P<0.01 indicate significant differences from control.

Table 2 Intracellular cyclic GMP levels measured in rat thoracic aorta under different experimental conditions

	cyclic GMP (pmol mg <sup>-1</sup> protein)	
Phenylephrine (0.3 $\mu$ M)	$9.98 \pm 1.49$	12
Acetylcholine (1 $\mu$ M)	$21.38 \pm 2.81**$	10
SR 58611 (30 μM)	$16.94 \pm 2.13**$	12
L-NMMA (100 $\mu$ M)	$8.81 \pm 1.48$	5
LNMMA + SR 58611	$8.91 \pm 1.01$	9
Denuded rings	$6.40 \pm 1.79$	5
Denuded rings + SR 58611	$11.95 \pm 2.23$	5

For each experimental condition, aortic rings were precontracted with phenylephrine in the presence or absence of L-NMMA or endothelium. In some rings, acetylcholine or cumulative concentrations of SR 58611 were added. At the end of the experiments, rings were frozen in order to measure intracellular cyclic GMP levels. Results show mean  $\pm$  s.e.mean. \*\*P<0.01 versus phenylephrine.

was also reduced (39.3  $\pm$  5.8% at 10  $\mu$ M isoprenaline; P < 0.01; Figure 4).

Relaxant effects of the  $\beta_3$ -adrenoceptor agonists, SR 58611, and CGP 12177

As shown in Figures 1C and 5, SR 58611 (0.1 – 30  $\mu$ M) induced concentration-dependent relaxation of rat aortic rings contracted with phenylephrine (0.3  $\mu$ M). This effect was similar to that observed with isoprenaline in the presence of nadolol. The relaxant effect of SR 58611 was initially slow, and maximal relevant response was achieved within 10-12 min (Figure 1C). As for the experiments in which isoprenaline was administrated after nadolol, the spontaneous relaxation obtained in control rings  $(18.1 \pm 5.2\%)$  relaxant effect at the end of the experiment; n=8) was subtracted from the corresponding relaxant effect of the  $\beta_3$ -adrenoceptor agonist. In these conditions, the pD<sub>2</sub> value was  $5.24 \pm 0.07$  (n = 11) and the E<sub>max</sub> value  $59.5 \pm 3.7\%$  (n=11) at a concentration of 30  $\mu$ M SR 58611 (Figure 5). Similar results were obtained with another contractile agent,  $PGF_{2\alpha}$  (3  $\mu M$ ) (data not shown). CGP 12177, a partial  $\beta_3$ -adrenoceptor agonist which also possesses  $\beta_1$ - and  $\beta_2$ -adrenoceptor antagonistic properties (Blin *et al.*, 1993), produced similar concentration-dependent relaxation. The pD<sub>2</sub> value was  $5.15\pm0.03$  (n=6) and the E<sub>max</sub> value  $63.9\pm5.2\%$ (n=6) at a concentration of 30  $\mu$ M CGP 12177. To analyse whether SR 58611 produced relaxant effects through activation of  $\beta_3$ -adrenoceptors alone, concentration response curves for this agonist were also determined in the presence of the  $\beta$ adrenoceptor antagonists, nadolol and SR 59230A (a  $\beta_3$ adrenoceptor antagonist). The concentration-response curve to SR 58611 was not modified by 30 min pre-treatment with 10  $\mu$ M nadolol (pD<sub>2</sub>=5.37±0.14; E<sub>max</sub>=50.6±3.3%; n=7) (Figure 6), whereas the effects of SR 58611 were abolished in the presence of SR 59230A (10  $\mu$ M; n=6; P<0.01 versus SR 58611 alone; Figure 6).

To characterize the involvement of the endothelium and of the NO synthase pathway in the vasorelaxant effect of  $\beta_3$ -adrenoceptor stimulation, the effect of endothelium removal and L-NMMA on the ability to modify contractile response to SR 58611 was determined. The relaxant effect of SR 58611 was shifted to the right after endothelium removal ( $E_{max} = 15.9 \pm 1.9\%$ ; pD<sub>2</sub>=4.88±0.03; n=7; P<0.01 versus SR 58611 alone; Figure 7). The relaxant effect of SR 58611 was also markedly attenuated by 30 min pre-treatment with 100  $\mu$ M L-NMMA (11.5±1.8% at 10  $\mu$ M SR 58611; n=9;

P < 0.01 versus SR 58611 alone); the pD<sub>2</sub> value was  $4.89 \pm 0.02$ (n=9). In these conditions, a relaxant effect of SR 58611 was only observed at the higher concentration of 30  $\mu$ M  $(27.0 \pm 2.5\%; n=9)$  (Figure 7).

## Intracellular cyclic GMP content

Intracellular cyclic GMP levels were measured in rat thoracic aorta rings at the end of the relaxation experiments in control and denuded rings pre-contracted with phenylephrine. As shown in Table 2, basal cyclic GMP levels were slightly lower in denuded rings and after 30 min pre-treatment with L-NMMA than in control rings. In rings with endothelium, cyclic GMP levels were increased 2 and 1.7 fold in the presence of acetylcholine (1 µM) and SR 58611 (30 µM) respectively. The SR 58611-induced increase in cyclic GMP levels was reduced by pre-treatment of aortic rings with L-NMMA (100  $\mu$ M) or after removal of the endothelium.

## Discussion

Several lines of evidence in this study show that activation of  $\beta_3$ -adrenoceptors significantly contributes to the relaxant effect of isoprenaline in rat thoracic aorta through activation of an endothelium-dependent NO synthase pathway. The relaxant effect of isoprenaline was still apparent after blockade of both  $\beta_1$ - and  $\beta_2$ -adrenoceptors by nadolol, and was similar to that exhibited both by the preferential  $\beta_3$ -adrenoceptor agonist, SR 58611, and by the partial  $\beta_3$ -adrenoceptor agonist, CGP 12177. The relaxant effect of SR 58611 was abolished by SR 59230A (a  $\beta_3$ -adrenoceptor antagonist) and greatly reduced by endothelium removal or by previous administration of L-NMMA. Finally, the increase in intracellular cyclic GMP levels observed in the same preparations after addition of SR 58611 was also abolished after previous administration of L-NMMA or endothelium removal.

Isoprenaline, a non-selective  $\beta$ -adrenoceptor agonist, induced rapid, concentration-dependent relaxation of rat thoracic aorta. Surprisingly, this effect persisted in the presence of nadolol, a potent  $\beta_1$ - and  $\beta_2$ -adrenoceptor antagonist with low affinity for native and recombinant  $\beta_3$ -adrenoceptors (Bond & Clarke, 1987; Emorine et al., 1989; Galitzky et al., 1993), but was shifted to the right with a reduction in the maximum. It is unlikely that this response was related to the competitive antagonism by nadolol, since the relaxation in the presence of nadolol was quite different from that obtained with isoprenaline alone. Thus, the relaxation developed more slowly and took a longer time to reach a steady-state than with isoprenaline alone. Thus, this relaxant effect may have been due to the stimulation of a third  $\beta$ -adrenoceptor subtype, possibly  $\beta_3$ , as suggested by our experiments conducted with two  $\beta_3$ adrenoceptor agonists, SR 58611 and CGP 12177. The relaxant effect of these two agents was similar to the slowly developing relaxations induced by another  $\beta_3$ -adrenoceptor agonist, BRL 37344, in isolated common carotid arteries of the rat (Oriowo, 1994) or smooth muscle of the gastrointestinal tract (McLaughlin & MacDonald, 1990; 1991). The potency of SR 58611 and CGP 12177 in rat thoracic aorta was similar to that of BRL 37344 and CGP 12177 in rat isolated carotid arteries (Oriowo, 1994), but much lower than previous values reported for the rat gastrointestinal tract (MacLaughlin & MacDonald, 1991; De Boer et al., 1993). It is still unclear why  $\beta_3$ adrenoceptor agonists have a poorer potency in vessels than in the gastrointestinal tract. In our study, the most convincing pharmacological evidence for the presence of  $\beta_3$ -adrenoceptors

in rat thoracic aorta was provided by  $\beta$ -adrenoceptor antagonists. The relaxant effect of SR 58611 was not modified by pretreatment with nadolol, indicating that this effect was not mediated by  $\beta_1$ - or  $\beta_2$ -adrenoceptors. This was confirmed by the relaxant effects achieved with CGP 12177, another  $\beta_1$ - or  $\beta_2$ adrenoceptor antagonist. Conversely, SR 59230A, a  $\beta_3$ adrenoceptor antagonist (Manara et al., 1995a; 1996), abolished the relaxant effect of SR 58611. To exclude a non selective effect of SR 59230 at the relatively high concentration used in our study, such as an activation of  $\beta_3$ -adrenoceptors (Strosberg & Pietri-Rouxel, 1996), we have performed some preliminary experiments with a lower concentration (1  $\mu$ M) of this antagonist. In these experiments, the inhibition of SR 58611-induced relaxation by 1  $\mu$ M SR 59230 persisted but was not as important as at 10 um SR 59230 (data not shown). In rat pulmonary vessels, several  $\beta_3$ -adrenoceptor agonists (SR 58611, SR 59119 and SR 59104) have produced relaxant effects. However, only the effect of SR 59104 was antagonized by SR 59230A (Dumas et al., 1998), which suggests that the pharmacology of vascular atypical  $\beta$ -adrenoceptors is complex.

The involvement of  $\beta_3$ -adrenoceptors in the relaxation of rat thoracic aorta and carotid arteries is consistent with the effects of  $\beta_3$ -adrenoceptor stimulation reported in vivo in dogs and rats.  $\beta_3$ -adrenoceptor stimulation produced peripheral vasodilatation, primarily in skin and adipose tissues in unanaesthetized dogs (Berlan et al., 1994; Shen et al., 1994), and to a lesser extent in rats (Shen et al., 1996). In dogs, studies using radioactive microspheres showed a different pattern of regional blood flow distribution after  $\beta_3$ -adrenoceptor stimulation as compared with isoprenaline-induced  $\beta_1$ - and  $\beta_2$ adrenoceptor-mediated peripheral vasodilatation. 37344, a  $\beta_3$ -adrenoceptor agonist administered in the presence of a  $\beta_1$ - and  $\beta_2$ -adrenoceptor antagonist, selectively increased blood flows in skin and adipose tissues (Shen et al., 1994). Vasodilatation has also been observed in brown adipose tissue of anaesthetized rats after administration of BRL 26830A, another  $\beta_3$ -adrenoceptor agonist, in the presence of arotinolol, a mixed  $\alpha$ -,  $\beta_1$ - and  $\beta_2$ -adrenoceptor antagonist (Takahashi et al., 1992). However,  $\beta_3$ -adrenoceptor agonists failed to induce vasodilatation in non-human primates (Shen et al., 1996).

In rat thoracic aortic rings, isoprenaline-induced relaxation persisted after endothelium removal, but only at higher concentrations and with a reduced maximal effect (Figure 2). In these conditions, addition of nadolol (10  $\mu$ M) abolished the relaxant effect of isoprenaline (data not shown). Similarly,  $\beta_3$ -adrenoceptor-induced relaxation was strongly reduced by endothelium removal, which suggests that  $\beta_3$ adrenoceptors are mainly located on endothelial cells. However, in isolated rat carotid arteries, the relaxation induced by two other  $\beta_3$ -adrenoceptor agonists (BRL 37344) and CGP 12177) was found to be endothelium-independent (Oriowo, 1994). Several factors could account for this discrepancy: (1) the vascular bed investigated; as previously described in the involvement of endothelium in  $\beta_1$ - and/or  $\beta_2$ -adrenoceptor mediated-relaxation (Gardiner *et al.*, 1991; Gray & Marshall, 1992; Graves & Poston, 1993; Béa et al., 1994); (2) the contractile agent used, i.e. norepinephrine in experiments with carotid arteries (Oriowo, 1994) and phenylephrine in the present study; and (3) the action of  $\beta_3$ -adrenoceptor agonists on  $\beta_1$ - and/or  $\beta_2$ -adrenoceptors. BRL 37344 activates  $\beta_1$ - and  $\beta_2$ -adrenoceptors, as well as  $\beta_3$ adrenoceptors, at high concentrations (Muzzin et al., 1992; Ida et al., 1996), and CGP 12177 may also stimulate the putative  $\beta_4$ -adrenoceptors previously described in the heart (Kaumann, 1996; 1997) and adipose tissue (Galitzky et al., 1997). Unfortunately, no monoclonal antibody selectively directed against  $\beta_3$ -adrenoceptors is currently available to confirm the location of these receptors on endothelial cells.

Pre-treatment of rat thoracic aorta with L-NMMA partially inhibited isoprenaline-induced relaxation, and the addition of nadolol produced a further rightward shift in the concentration-response curve of the drug. SR 58611 also produced a slight relaxation in the presence of L-NMMA. These results strongly suggest that  $\beta_3$ -adrenoceptors mediate vascular relaxation through the activation of an NO synthasedependent pathway. Conversely, both  $\beta_1$  and  $\beta_2$ -adrenoceptors relax rat thoracic aorta through activation of an NOdependent (Gray & Marshall, 1992; Wang et al., 1993) and/ or -independent (Eckly et al., 1994) pathway.  $\beta_3$ -adrenoceptorinduced relaxation was associated with an increase in intracellular cyclic GMP levels, which was abolished by pretreatment with L-NMMA. Thus, the activation of vascular  $\beta_3$ adrenoceptors stimulated the NO synthase pathway, leading to an increase in intracellular cyclic GMP levels. An NO synthase pathway has also been described in human ventricle recently (Gauthier et al., 1998) in which  $\beta_3$ -adrenoceptor stimulation induced a negative inotropic effect secondary to increases in both NO production and cyclic GMP levels. These effects were blunted after NO synthase inhibition (Gauthier et al., 1998). Thus, the signalling pathway of  $\beta_3$ -adrenoceptors in both heart and vessels seems to be different from that described in adipose tissues where  $\beta_3$ -adrenoceptors, like  $\beta_1$ - and  $\beta_2$ -adrenoceptors,

stimulate a cyclic AMP pathway (for review, see Strosberg, 1997).

The characterization of a  $\beta_3$ -adrenoceptor subtype in vessels, in addition to  $\beta_1$  and  $\beta_2$ , raises the question of the role of these  $\beta_3$ -adrenoceptors in vasorelaxation. As the stimulation of these receptors produces vasorelaxation, as in the case with  $\beta_2$ -adrenoceptor stimulation,  $\beta_3$ -adrenoceptors could play a redundant function in vessels in which  $\beta_2$ - and  $\beta_3$ adrenoceptors are co-expressed. It has been shown that catecholamines stimulate  $\beta_3$ -adrenoceptors in rat and dog fat cells at higher concentrations than those required to recruit  $\beta_1$ and  $\beta_2$ -adrenoceptors (Granneman, 1992; Galitzky et al., 1993). Furthermore,  $\beta_3$ -adrenoceptors, unlike  $\beta_1$ - and  $\beta_2$ subtypes, lack regulatory phosphorylation sites for G protein receptor kinases (Liggett et al., 1993) and could be relatively resistant to agonist-induced desensitization. Thus,  $\beta_3$ -adrenoceptors may be involved when the sympathetic nervous system is overstimulated.

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#### References

- BEA, M.L., GHALEH, B., GIUDICELLI, J.F. & BERDEAUX, A. (1994). Lack of importance of NO in β-adrenoceptor-mediated relaxation of large epicardial canine coronary arteries. *Br. J. Pharmacol.*, **111**, 981–982.
- BERLAN, M., GALITZKY, J., BOUSQUET-MELOU, A., LAFONTAN, M. & MONTASTRUC, J.L. (1994). *Beta-3* adrenoceptor-mediated increase in cutaneous blood flow in the dog. *J. Pharmacol. Exp. Ther.*, **268**, 1444–1451.
- BLIN, N., CAMOIN, L., MAIGRET, B. & STROSBERG, A.D. (1993). Structural and conformational features determining selective signal transduction in the beta 3-adrenergic receptor. *Mol. Pharmacol.*, **44**, 1094–1104.
- BOND, R.A. & CLARKE, D.E. (1987). A response to isoprenaline unrelated to alpha and beta-adrenoceptor antagonism. *Br. J. Pharmacol.*, **91**, 683–686.
- CLARK, B.J. & BERTHOLET, A. (1983). Effects of pindolol on vascular smooth muscle. *Gen. Pharmacol.*, 14, 117-119.
- DE BOER, R.E., BROUWER, F. & ZAAGSMA, J. (1993). The beta-adrenoceptors mediating relaxation of rat oesophageal muscularis mucosae are predominantly of the beta 3-, but also of the beta 2-subtype. *Br. J. Pharmacol.*, **110**, 442–446.
- DOGGRELL, S.A. (1990). Relaxant and beta 2-adrenoceptor blocking activities of (+/-)-, (+)- and (-)-pindolol on the rat isolated aorta. *J. Pharm. Pharmacol.*, **42**, 444-446.
- DUMAS, M., DUMAS, J.P., BARDOU, M., ROCHETTE, L., ADVENIER, C. & GIUDICELLI, J.F. (1998). Influence of β-adrenoceptor agonists on the pulmonary circulation. Effects of a β<sub>3</sub>-adrenoceptor antagonist, SR 59230A. *Eur. J. Pharmacol.*, **348**, 223–228.
- ECKLY, A.E., STOCLET, J.C. & LUGNIER, C. (1994). Isoprenaline induces endothelium-independent relaxation and accumulation of cyclic nucleotides in the rat aorta. *Eur. J. Pharmacol.*, **271**, 237–240.
- EMORINE, L.J., MARULLO, S., BRIEND-SUTREN, M.M., PATEY, G., TATE, K., DELAVIER-KLUTCHKO, C. & STROSBERG, A.D. (1989). Molecular characterization of the human  $\beta_3$ -adrenergic receptor. *Sciences*, **245**, 1118–1121.

- GALITZKY, J., LANGIN, D., VERWAERDE, P., MONTASTRUC, J.L., LAFONTAN, M. & BERLAN, M. (1997). Lipolytic effects of conventional beta 3-adrenoceptor agonists and of CGP 12,177 in rat and human fat cells: preliminary pharmacological evidence for a putative beta 4-adrenoceptor. Br. J. Pharmacol., 122, 1244– 1250.
- GALITZKY, J., REVERTE, M., CARPENE, C., LAFONTAN, M. & BERLAN, M. (1993). Beta3-adrenoceptors in dog adipose tissue: studies on their involvement in the lipomobilizing effect of catecholamines. *J. Pharmacol. Exp. Ther.*, **266**, 358 366.
- GARDINER, S.M., KEMP, P.A. & BENNETT, T. (1991). Effects of N<sup>G</sup>-nitro-L-arginine methyl ester on vasodilator responses to acetylcholine, 5'-N-ethylcarboxamidoadenosine or salbutamol in conscious rats. *Br. J. Pharmacol.*, **103**, 1725–1732.
- GAUTHIER, C., LEBLAIS, V., KOBZIK, L., TROCHU, J.N., KHANDOUDI, N., BRIL, A., BALLIGAND, J.L. & H. LE MAREC. (1998). The negative inotropic effect of  $\beta_3$ -adrenoceptor stimulation is mediated by activation of a nitric oxide synthase pathway in human ventricle. *J. Clin. Invest.*, **102**, 1377–1384.
- GAUTHIER, C., TAVERNIER, G., CHARPENTIER, F., LANGIN, D. & H. LE MAREC. (1996). Functional  $\beta_3$ -adrenoceptor in the human heart. *J. Clin. Invest.*, **98**, 556–562.
- GRANNEMAN, J.G. (1992). Effects of agonist exposure on the coupling of beta1- and beta3-adrenergic receptors to adenylyl cyclase in isolated adipocytes. *J. Pharmacol. Exp. Ther.*, **261**, 638-642.
- GRAVES, J. & POSTON, L. (1993). Beta-adrenoceptor agonist mediated relaxation of rat isolated resistance arteries: a role for the endothelium and nitric oxide. *Br. J. Pharmacol.*, **108**, 631–637.
- GRAY, D.W. & MARSHALL, I. (1992). Novel signal transduction pathway mediating endothelium-dependent β-adrenoceptor vasorelaxation in rat thoracic aorta. Br. J. Pharmacol., 107, 684– 690.

- IDA, K., HASHIMOTO, K., KAMIYA, M., MUTO, S., NAKAMURA, Y., KATO, K. & MIZOTA, M. (1996). Stereoselective action of  $(R^*,R^*)$ -(+)-methyl-4-[2[2-hydroxy-2(3-chlorophenyl)ethylamino[propyl]-phenoxyacetic acid (BRL 37344) on  $\beta$ -adrenoceptors and metabolic chiral inversion. Biochem. Pharmacol., 52, 1521 – 1527.
- KAUMANN, A.J. (1996). (-)-CGP 12177-induced increase of human atrial contraction through a putative third  $\beta$ -adrenoceptor. Br. J. *Pharmacol.*, **117**, 93–98.
- KAUMANN, A.J. (1997). Four beta-adrenoceptor subtypes in the mammalian heart. Trends Pharmacol. Sci., 18, 70-76
- LAFONTAN, M. (1994). Differential recruitment and differential regulation by physiological amines of fat cell  $\beta_1$ -,  $\beta_2$ - and  $\beta_3$ adrenergic receptors expressed in native fat cells and in transfected cell lines. *Cell Signalling*, **4**, 363–392.
- LANDS, A.M., ARNOLD, A., MCAULIFF, J.P., LUDUENA, F.P. & BROWN, T.G. (1967). Differentiation of receptor systems activated by sympathomimetic amines. *Nature*, **214**, 597 – 598.
- LEE, R.J., EVANS, D.B., BAKY, S.H. & LAFFAN, R.J. (1975). Pharmacology of nadolol (SQ 11725), a beta-adrenergic antagonist lacking direct myocardial depression. Eur. J. Pharmacol., 33, 371 – 382.
- LIGGETT, S.B., FREEDMAN, N.J., SCHWINN, D.A. & LEFKOWITZ, R.J. (1993). Structural basis for receptor subtype-specific regulation revealed by a chimeric  $\beta_3$ -/ $\beta_2$ -adrenergic receptor. Proc. Natl. Acad. Sci. U.S.A., 90, 3665-3669.
- LOWRY, O.H., ROSEBROUGH, N.J., FARR, A.L. & RANDALL, R.J. (1951). Protein measurement with the Folin phenol reagent. J. Biol. Chem., 193, 265-275.
- MANARA, L., BADONE, D., BARONI, M., BOCCARDI, G., CECCHI, R., CROCI, T., GIUDICE, A., GUZZI, U., LANDI, M. & LE FUR, G. (1996). Functional identification of rat atypical  $\beta$ -adrenoceptors by the first  $\beta_3$ -selective antagonists, aryloxypropanolaminotetralins. Br. J. Pharmacol., 117, 435-442.
- MANARA, L., BADONE, D., BARONI, M., BOCCARDI, G., CECCHI, R., CROCI, T., GIUDICE, A., GUZZI, U. & LE FUR, G. (1995a). Aryloxypropanolaminotetralins are the first selective antagonists for atypical  $(\beta_3)$   $\beta$ -adrenoceptors. *Pharmacol. Commun.*, **6**, 253 –
- MANARA, L., CROCI, T. & LANDI, M. (1995b).  $\beta_3$ -adrenoceptors and intestinal motility. Fundam. Clin. Pharmacol., 9, 332-342.
- MARTIN, C.A.E. & ADVENIER, C. (1995a). Beta 3-adrenoceptors and airways. Fundam. Clin. Pharmacol., 9, 114-118.
- MCLAUGHLIN, D.P. & MACDONALD, A. (1990). Evidence for the existence of 'atypical' beta-adrenoceptors (beta 3-adrenoceptors) mediating relaxation in the rat distal colon in vitro. Br. J. Pharmacol., 101, 569 – 574.

- MCLAUGHLIN, D.P. & MACDONALD, A. (1991). Characterization of catecholamine-mediated relaxations in rat isolated gastric fundus: evidence for an atypical beta-adrenoceptor. Br. J. Pharmacol., 103, 1351-1356.
- MUZZIN, P., REVELLI, J.P., FRASER, C.M. & GIACOBINO, J.P. (1992). Radioligand binding studies of the atypical  $\beta_3$ -adrenergic receptor in rat brown adipose tissue using [3H]CGP 12177. FEBS Lett., 298, 162-164.
- O'DONNELL, S.R. & WANSTALL, J.C. (1985). Responses to the beta 2-selective agonist procaterol of vascular and atrial preparations with different functional beta-adrenoceptor populations. Br. J. Pharmacol., 84, 227-235.
- ORIOWO, M.A. (1994). Atypical  $\beta$ -adrenoceptors in the rat isolated common carotid artery. Br. J. Pharmacol., 113, 699-702.
- ORIOWO, M.A. (1995). Different atypical beta-adrenoceptors mediate isoprenaline-induced relaxation in vascular and nonvascular smooth muscles. Life Sci., 56, PL269-PL275.
- SHEN, Y.T., CERVONI, P., CLAUS, T. & VATNER, S.F. (1996). Differences in beta 3-adrenergic receptor cardiovascular regulation in conscious primates, rats and dogs. J. Pharmacol. Exp. Ther., 278,  $1435 - \overline{1443}$ .
- SHEN, Y.T., ZHANG, H. & VATNER, S.F. (1994). Peripheral vascular effects of Beta-3 adrenergic receptor stimulation in conscious dogs. J. Pharmacol. Exp. Ther., 268, 466-473.
- STROSBERG, A.D. (1997). Structure and function of the  $\beta_3$ adrenergic receptor. Annu. Rev. Pharmacol. Toxicol., 37, 421-
- STROSBERG, A.D. & PIETRI-ROUXEL, F. (1996). Function and regulation of the beta 3-adrenoceptor. Trends Pharmacol. Sci., **17,** 373 – 381
- TAKAHASHI, H., YOSHIDA, T., NISHIMURA, M., NAKANISHI, T., KONDO, M. & YOSHIMURA, M. (1992). Beta3-adrenergic agonist, BRL-26830A, and alpha/beta blocker, arotinolol, markedly increase regional blood flow in the brown adipose tissue in anesthetized rats. Jpn. Circ. J., 56, 936-942.
- TAVERNIER, G., GALITZKY, J., BOUSQUET-MELOU, A., MONTAS-TRUC, J.L. & BERLAN, M. (1992). The positive chronotropic effect induced by BRL 37344 and CGP 12177, two beta-3 adrenergic agonists, does not involve cardiac beta adrenoceptors but baroreflex mechanisms. J. Pharmacol. Exp. Ther., 263, 1083-1090.
- WANG, Y.X., POON, K.S., RANDALL, D.J. & PANG, C.C.Y. (1993). Endothelium-derivated nitric oxide partially mediates salbutamol-induced vasodilatations. Eur. J. Pharmacol., 250, 335-340.

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